Comparison between two automated chemiluminescence immunoassays for quantifying 25 (OH) vitamin D

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Summary

Introduction: Quantifying total blood 25 (OH) vitamin D is the most accurate marker of an individual’s vitamin D status. The gold standard technique for measurement is liquid chromatography tandem mass spectrometry (LC-MS/MS), although currently clinical laboratories tend to use chemiluminescence techniques. The objective of this study was to compare 25 (OH) vitamin D concentrations obtained by two commercially-produced automated methods and study the correlation of these methods with the LC-MS/MS reference technique.

Material and methods: The 25(OH) vitamin D levels were quantified in 1,000 serum samples from the Jimenez Diaz Biochemistry Foundation Laboratory using 2 automated methods for chemiluminescence detection: ADVIA CENTAUR® (SIEMENS) and LUMIPULSE® G1200 (Fujirebio). Among all the samples tested, the 50 most discordant to each other were sent to be evaluated by LC-MS/MS reference technique.

Results: The results indicate that there is good correlation between the two methods: CCI=0.923 (0.914-0.932), with the G1200 LUMIPULSE® values 10% being higher than CENTAUR®. Regarding the 50 samples selected, we can see that there is a good correlation between the two immunoassays with LC-MS/MS, although both methods significantly underestimate 25 (OH) vitamin D results with respect to the gold standard.

Discussion: Although both techniques are suitable for use, it is worth considering whether the worldwide vitamin D deficiency epidemic is due to the analysis methodology used. This variability between immunoassays could be solved by standardizing the different commercial techniques in line with NIST-produced reference materials.

Keywords: 25(OH) vitamin D, Fujirebio, SIEMENS, technical comparison, LC-MS/MS.
Introduction

Vitamin D is a fat-soluble vitamin involved in calcium and phosphorus metabolism whose role is essential in bone formation and mineralization. Currently, its immunomodulatory actions, antiproliferative and cell differentiation stimulatory of associated pathologies such as cardiovascular diseases, diabetes and cancer have also been shown1,2.

Quantification of 25 (OH) vitamin D total blood is the most accurate marker of vitamin D status in an individual, although its active metabolite is 1,25 (OH) 2 vitamin D3,4.

The gold standard technique for measurement is liquid chromatography/tandem mass spectrometry (LC-MS/MS), but clinical laboratories routinely used chemiluminescence immunoassays5.

The main problems of these immunoassays are due to the hydrophobic nature of the analyte, the high concentration in which vitamin D binding protein (DBP) in serum is found, and the existence of cross reactions of multiple vitamin D with the metabolite antibodies used in the process. Therefore, proper immunoassay requires a pretreatment to inactivate DBP, a careful selection of antibody used and the standardization of the technique compared to the values returned by LC-MS/MS analyzer. Different commercial techniques for analysis of vitamin D differ in how to separate the binding protein, the percentage of crossed reactions with other metabolites of our analyte, as well as the specificity of the antibody used6.

Currently, as a result of general knowledge on severe vitamin D deficiency in the world's population, it has become necessary to measure vitamin D levels in different populations, research cohorts and individual patient7. Several studies have shown considerable variation between the different analytical methods based on immunochrometry, liquid/UV and LC-MS/MS chromatography. It has been argued that a particular patient can be classified with levels of sufficiency or insufficiency of vitamin D depending on the laboratory where this analysis is carried out8.

To solve this problem, the need for standardization of the levels of 25 (OH) vitamin D has been established by numerous scientific organizations. In 2011, the Office of Dietary Supplements (ODS) of the National Institutes of Health US (NIH) in collaboration with the National Institute of Standards and Technology (NIST) created the program standardization of vitamin D (VDSP). NIST has developed 4 based reference materials with different serum concentrations of vitamin D known order to standardize the various commercial techniques9. However, not all the current methods used to quantify 25(OH) vitamin D already calibrated against these accepted standards10.

The aim of this study was to compare 25(OH) vitamin D concentrations obtained using two commercial automated methods and study the correlation of these methods with the LC-MS/MS technique.

Material and methods

We used 1,000 serum samples selected at random from those tested in FJD’s clinical analysis laboratory. The samples were obtained from patients aged between 1 and 92 years (59±18, average ± SD) with 37% of women and 63% men. Levels of 25 (OH) vitamin D were quantified with ADVIA Centaur XP® (SIEMENS) and LUMIPULSE® G1200 (FUJI REBIO). In all cases the samples were handled anonymously, so obtaining informed consent of patients was not required.

The analyzer LUMIPULSE® G1200 (FUJI REBIO) takes a noncompetitive sandwich-type immunoassay with chemiluminescent detection using two antibodies, a monoclonal sheep antibody that binds to 25(OH) vitamin D2 and D3, and a second monoclonal antibody that binds exclusively to the complex formed above. Separation of vitamin D binding protein is carried out by a chemical agent in 1st reaction.

According to the manufacturer's specifications, the technique has an intra-assay imprecision with a coefficient of variation (CV) ≤6% and a functional sensitivity of 3.491 ng/mL. Measurement interval is 4-150 ng/mL. Analytical specificity reflected by the percentage of cross-reactivity with other metabolites is 100% for 25(OH) vitamin D3, 100.1% for 25(OH) vitamin D2, and 19.9% for the epimer C3 25(OH) vitamin D3.

The ADVIA Centaur XP® (SIEMENS) is a competitive immunoassay with chemiluminescent detection using an anti-fluorescein murine monoclonal antibody covalently coupled to paramagnetic particles (PMP), a murine monoclonal anti-25(OH) vitamin D marked with acridinium ester, and vitamin D analog with fluorescein. As separation medium binding protein release agent is used in buffered saline.

According to the manufacturer's specifications, the technique presents an intra-assay imprecision with a CV of 4.2% -11.9% and a functional sensitivity of 4.2 ng/mL. Measurement interval is 4.2 to 150 ng/mL. Analytical specificity reflected by the percentage of cross-reactivity with other metabolites is 97.4% for the 25(OH) vitamin D3, from 106.2% for 25 (OH) vitamin D2 and 1% for C3 epimer 25(OH) vitamin D3.

Among all samples tested, the 50 discordant with each other were sent to be evaluated by LC-MS/MS method in the laboratory of Dr. Etienne Cavalier (Department of Clinical Chemistry, University of Liege, Belgium); in order to compare the two chemiluminescent immunoassays relative to the reference technique LC-MS/MS. The difference between these results 50 samples ranged between 14% and 133% (32±52%, mean ± SD) with respect to the average of the two values obtained. In all cases the percentage was higher than the coefficients of variation inter-analysis of the two methods: FUJI REBIO, 6%; SIEMENS, 11.9%. We analyzed the most discordant samples to ascertain whether they belonged to a particular group of patients. For example, pregnant women who have abnormal levels of vitamin D binding
protein. However, we note that these patients belonged mainly to Nephrology, Rheumatology and Endocrinology, an insignificant fact since these services are most demanded the determination of vitamin D. Moreover, the average age of these 50 patients was 63±16 years, with 34% male and 66% female, figures which were quite similar to the total group of 1,000 samples (age 59±18 years with 37% of women and 63% men). The most discordant samples were chosen together to analyze for gas-mass because our goal was two-fold. First, to check its similarity to the technique of gas-mass and otherwise clarify which of the two techniques was closer to the reference technique. This second point could not clarify if we sent the samples whose results were similar. Regarding the choice of 50 samples as appropriate for the study was done because it was a sufficient amount to obtain statistically significant results samples. Due to the high cost of gas-mass determination it was not possible to send a larger number of samples.

Results

We assessed the degree of concordance of vitamin D measures provided by the two appliances: ADVIA Centaur XP® and LUMIPULSE G1200®. To do this, we calculated the intraclass correlation coefficients (ICC) with confidence intervals at 95%. The results indicate a good correlation between the two methods: CCI=0.923 (0.914 to 0.932). There are no significant differences in the ICC if the samples are divided into groups with vitamin D levels \(\leq 20\) ng/mL and > 20 ng/mL.

The regression line obtained from both trials was \(Y=1.221+1.035X\), where \(Y\) corresponds to the values of LUMIPULSE G1200 and \(X\) to Centaur XP®. LUMIPULSE G1200 values were 10% higher than Centaur® (Figure 1).

With respect to 50 samples selected subgroup to analyze by LC-MS/MS was obtained with ICC=0.987 LUMIPULSE and CCI=0.938 with Centaur®. Although both are satisfactory, the intraclass correlation coefficient is the highest LUMIPULSE therefore measurements of this device is more like the exact ones (Figures 2 and 3).

Then with our subgroup of 50 selected samples, we carried out the Bland-Altman plots, where the X axis corresponds to the mean of each pair of observations and the Y axis differences between each pair of observations. In the graphs there are two horizontal solid lines. The gray solid line is drawn at zero height; if the measurements given by the apparatus were identical to the exact measurement points should be located right on this line. The solid blue line represents the mean of the differences. If this line is the line below the 0 value means that the device tends to lower measures the exact value, and if it is above the opposite.

As can be seen in figure 4, the mean difference between the analyzer LUMIPULSE® G1200 and the reference method LC-MS/MS is 20%; therefore this immunoassay underestimates values of 25 (OH) vitamin D by 20% compared to the gold standard. In figure 5, we see how in the case of the Centaur® the average of the differences is 42%, so the values which casts this technique are much lower than those of the reference method. Furthermore, the technique LUMIPULSE® G1200 is less spread in the results.

Discussion

Both methods have a good correlation between them, the values obtained in the Centaur® approximately 10% lower than those obtained by the LUMIPULSE® G1200.
The correlation of both immunoassays with the reference technique LC-MS/MS is good (although higher for LUMIPULSE than Centaur), which does not exclude that the two methods considerably underestimate the results of 25(OH) vitamin D with respect to gold standard. In this selection of samples, held by choosing those where the discrepancy between the two methods was higher, LUMIPULSE® G1200 underestimates the values by 20%, while yields Centaur® 25 (OH) vitamin D 42% lower than the reference method LC-MS/MS. This difference between the two immunoassays is given by the different technique used (competitive assay in SIEMENS and noncompetitive sandwich in FUJIREBIO), pretreatment of the sample to separate the 25 (OH) vitamin D and DBP selected antibodies.

Currently, according to studies, more than half of the world's population has insufficient levels or even frank deficiency of vitamin D. This may be considered a global “epidemic”, but should be questioned as to whether this state of widespread vitamin deficiency is largely influenced by the analysis methodology used in determining concentrations of 25(OH) vitamin D8,11.

Figure 3. Regression line calculated by Passing-Bablok between Centaur® (SIEMENS) and LC-MS/MS using 50 selected samples (see Materials and methods)

Figure 4. Bland-Altman Charts between LUMIPULSE® G1200 (FUJIREBIO) and LC-MS/MS using 50 selected samples (see Materials and methods)

Figure 5. Bland-Altman Charts between Centaur® (SIEMENS) and LC-MS/MS using 50 selected samples (see Materials and methods)
This variability between immunoassays would be solved by standardizing different commercial techniques with reference materials for measuring 25(OH) vitamin D produced by the NIST.

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Bibliography